

Supplementary data to:

The evolutionary history of colour polymorphism in *Ischnura* damselflies (Odonata: Coenagrionidae)

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Odonatologica 49(3/4) 2020: 333-370 – DOI:10.5281/zenodo.4268559

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Methodological details. DNA extraction and sequencing

Previous phylogenetic studies included 15 Nearctic and Neotropical taxa (CHIPPINDALE et al. 1999) and 24 old-world species (mainly Palaearctic taxa; DUMONT 2013). We included 44 taxa in our study, and used two mitochondrial genes, cytochrome oxidase II (COII) and cytochrome b (CYTB), and a part of two nuclear genes, the small sub-unit 18S nrDNA (ITS1) and 5.8S (ITS2), and the large sub-unit rDNA 28 nrDNA (18S-ITS). For species

lacking published sequences (*Ischnura* sp. “a”, *I. chingaza*, *I. cruzi*, *I. cyane*, *I. foylei*, and *I. indivisa*), DNA was extracted from the thorax using a standard phenol/chloroform-isoamylalcohol extraction protocol (SAMBROOK et al. 1989). Table S1 shows the corresponding sequence accession numbers. DNA samples were amplified by PCR using universal primers: TL2-J-3037 and C2-N-3494 and C2-J-3400 and TK-N-3785 (n=673 bp) for COII (SIMON et al. 1994); CB-J-10933 and TS1-N-11683 (n=457 bp) for CYTB (SIMON et al. 1994); and: ITS-1 and reverse 28H (n=751 bp) for 18S-ITS (SAMRAOUI et al. 2002). DNA amplifications were done following the protocol described in SÁNCHEZ-GUILLÉN et al. (2014). Bidirectional sequencing was conducted, and the sequencing was completed on an ABI 3100 Genetic Analyzer. Because we have sequenced DNA from five dried specimens with partially degraded DNA, the amplification of the tree genes was not possible in the six species (cf. Table S1). Forward and reverse sequences were edited in GeneStudio v.2.2.0.0. Variable positions were revised by eye, and, based on the chromatogram, only high-quality sequences were considered. Sequenced data from old or dried specimens may be incomplete due to degraded DNA. Therefore, we only processed high-quality sequences for each gene and taxon and then selected the longest sequences of these to maximize data coverage (GenBank accession numbers, Table S1).

References

- CHIPPINDALE P.T., DAVE V.K., WHITMORE D.H. & ROBINSON J.V. 1999. Phylogenetic relationships of North American damselflies of the genus *Ischnura* (Odonata: Zygoptera: Coenagrionidae) based on sequences of three mitochondrial genes. *Molecular Phylogenetics and Evolution* 11: 110-121
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- SIMON C., FRATI F., BECKENBACH A., CRESP B., LIU H. & FLOOK P. 1994. Evolution, weighting, and phylogenetic utility of mitochondrial gene-sequences and a compilation of conserved polymerase chain-reaction primers. *Annals of the Entomological Society of America* 87: 651-701

Table S1. Sequence references and GenBank accession numbers. MORGAN (2002) – data from: MORGAN J.L. 2002. Worldwide phylogeny of the damselfly genus *Ischnura* based on mitochondrial cytochrome oxidase II and cytochrome B sequence data. Master thesis, The University of Texas at Arlington; na – data were not available.

Species	References		
	Gene COII	Gene CYTB	Gene ITS
<i>Ischnura</i> sp. "a"	na	MG561437	na
<i>Ischnura abyssinica</i>	na	na	FN356098.1
<i>Ischnura aralensis</i>	na	na	FN356099
<i>Ischnura asiatica</i>	KC430145	KC430194	AB706602
<i>Ischnura aurora</i>	MORGAN (2002)	MORGAN (2002)	AB706608
<i>Ischnura barberi</i>	AF067663	AF067683	na
<i>Ischnura capreolus</i>	AF067682	AF067684	na
<i>Ischnura cervula</i>	AF067665	AF067685	FN356101
<i>Ischnura chingaza</i>	na	MG561433	MG561438
<i>Ischnura cruzi</i>	na	MG561435	MG561439
<i>Ischnura cyane</i>	na	MG561434	MG561440
<i>Ischnura damula</i>	AF067666	AF067686	na
<i>Ischnura demorsa</i>	AF067667	AF067687	na
<i>Ischnura denticollis</i>	AF067668	AF067688	FN356102
<i>Ischnura elegans</i>	HQ834805	HQ834800	KC430216
<i>Ischnura e. ebneri</i>	JQ911513	KC430182	JQ911510
<i>Ischnura erratica</i>	AF067671	AF067691	na
<i>Ischnura evansi</i>	MORGAN (2002)	MORGAN (2002)	na
<i>Ischnura ezoin</i>	na	na	AB706574
<i>Ischnura fluviatilis</i>	MORGAN (2002)	MORGAN (2002)	na
<i>Ischnura foylei</i>	MG561432	na	na
<i>Ischnura forcipata</i>	na	na	FN356104.1
<i>Ischnura fountaineae</i>	KC430124	KC430174	KC430212
<i>Ischnura gemina</i>	AF067672	AF067692	na
<i>Ischnura genei</i>	KC430116	KC430152	KC430199
<i>Ischnura graellsii</i>	HQ834807	KC430158	KC430203
<i>Ischnura hastata</i>	AF067673	AF067693	na
<i>Ischnura heterosticta</i>	MORGAN (2002)	MORGAN (2002)	AB706612
<i>Ischnura indivisa</i>	na	MG561436	na

Species	References		
	Gene COII	Gene CYTB	Gene ITS
<i>Ischnura intermedia</i>	MORGAN (2002)	KY127445	na
<i>Ischnura kellicotti</i>	AF067675	AF067695	na
<i>Ischnura nursei</i>	na	na	MH447413
<i>Ischnura perparva</i>	AF067676	AF067696	FN356106
<i>Ischnura posita atezca</i>	AF067677	AF067697	na
<i>Ischnura p. posita</i>	na	AF067698	na
<i>Ischnura prognata</i>	AF067679	AF067699	na
<i>Ischnura pumilio</i>	KC430197	KC430198	KC430231
<i>Ischnura ramburii</i>	AF067680	AF067700	na
<i>Ischnura rubilio</i>	na	na	MH447434
<i>Ischnura rufostigma</i>	na	na	AB706613
<i>Ischnura saharensis</i>	KC430123	KC430168	KC430211
<i>Ischnura senegalensis</i>	KC430142	KC430189	KC430219
<i>Ischnura ultima</i>	MORGAN (2002)	MORGAN (2002)	na
<i>Ischnura verticalis</i>	AF067682	AF067702	na
<i>Enallagma basidens</i>	na	AF067689.1	na
<i>Enallagma civile</i>	na	AF067690.1	na

Table S2. Species distribution and biogeographic realms

Information was taken from the following multiple databases providing open access to biodiversity data:

GBIF – Global Biodiversity Information Facility.

Online on the Internet, URL (30-x-2020): <http://www.gbif.org/species/1423281>
Dragonfiles 0.9.

Online on the Internet, URL (30-x-2020): <http://www.medusa.jcu.edu.au>

COL – Catalogue of Life.

Online on the Internet, URL (30-x-2020): www.catalogueoflife.org

EOL – Encyclopedia of Life.

Online on the Internet, URL (30-x-2020): <https://eol.org/>

Species	Distribution range	Biogeographic realms
<i>Ischnura</i> sp. „a“	Colombia	Neotropical
<i>Ischnura abyssinica</i> Martin, 1908	Ethiopia	Afrotropical
<i>Ischnura acuticauda</i> Lieftinck, 1959	New Guinea	Australasian
<i>Ischnura albistigma</i> Fraser, 1927	Samoa	Australasian
<i>Ischnura aralensis</i> Haritonov, 1979	Kazakhstan, Russia, Uzbekistan	Palaeartic
<i>Ischnura ariel</i> Lieftinck, 1949	New Guinea	Australasian
<i>Ischnura armeniacum</i> Lieftinck, 1949	New Guinea	Australasian
<i>Ischnura asiatica</i> (Brauer, 1865)	East Asia	Palaeartic
<i>Ischnura aurora</i> (Brauer, 1865)	Asia, Australia, Pacific	Australasian-Indomalayan
<i>Ischnura barberi</i> Currie, 1903	SW North America, Mexico	Nearctic
<i>Ischnura buxtoni</i> Fraser, 1927	Samoa	Australasian
<i>Ischnura capreolus</i> (Hagen, 1861)	Central-South America	Neotropical
<i>Ischnura cardinalis</i> Kimmins, 1929	Society Islands (Polynesia)	Australasian
<i>Ischnura cervula</i> Selys, 1876	W North America	Nearctic
<i>Ischnura chingaza</i> Realpe, 2010	Colombia	Neotropical
<i>Ischnura chromostigma</i> Fraser, 1927	Samoa	Australasian
<i>Ischnura cruzi</i> De Marmels, 1987	Colombia	Neotropical
<i>Ischnura cyane</i> Realpe, 2010	Colombia	Neotropical
<i>Ischnura damula</i> Calvert, 1902	W North America	Nearctic
<i>Ischnura demorsa</i> (Hagen, 1861)	W North America	Nearctic
<i>Ischnura denticollis</i> (Burmeister, 1839)	W North America, Mexico	Nearctic
<i>Ischnura dorothea</i> Fraser, 1924	China	Palaeartic

Species	Distribution range	Biogeographic realms
<i>Ischnura elegans</i> (Vander Linden, 1820)	Eurasia	Palearctic
<i>Ischnura erratica</i> Calvert, 1895	W North America	Nearctic
<i>Ischnura evansi</i> Morton, 1919	Iran, Iraq	Palearctic
<i>Ischnura ezoin</i> (Asahina, 1952)	Ogasawara Islands (Japan)	Palearctic
<i>Ischnura filosa</i> Schmidt, 1951	Madagascar	Afrotropical
<i>Ischnura fluviatilis</i> Selys, 1876	South America	Neotropical
<i>Ischnura forcipata</i> Morton, 1907	Asia	Palearctic
<i>Ischnura fountaineae</i> Morton, 1905	North Africa, Pantelleria	Palearctic
<i>Ischnura foylei</i> Kosterin, 2015	Sumatra	Indomalayan
<i>Ischnura gemina</i> (Kennedy, 1917)	California	Nearctic
<i>Ischnura genei</i> (Rambur, 1842)	Tyrrhenian Islands, Sicily, Maltese Islands	Palearctic
<i>Ischnura graellsii</i> (Rambur, 1842)	Iberia, North Africa	Palearctic
<i>Ischnura haemastigma</i> Fraser, 1927	Samoa	Australasian
<i>Ischnura halecarpenteri</i> Mundford, 1942	Marquesas Islands (Polynesia)	Australasian
<i>Ischnura hastata</i> (Say, 1839)	America, Azores	Neotropical-Nearctic
<i>Ischnura heterosticta</i> (Burmeister, 1839)	Australia	Australasian
<i>Ischnura inarmata</i> Calvert, 1898	Kashmir	Palearctic
<i>Ischnura indivisa</i> (Ris, 1918)	Costa Rica	Neotropical
<i>Ischnura intermedia</i> Dumont, 1974	Turkey	Palearctic
<i>Ischnura isoetes</i> Lieftinck, 1949	New Guinea	Australasian
<i>Ischnura jeanyvesmeyeri</i> Englund & Polhemus, 2010	Polynesia	Australasian
<i>Ischnura karafutonis</i> Matsumura, 1931	Japan	Palearctic
<i>Ischnura kellicotti</i> Williamson, 1898	W North America	Nearctic
<i>Ischnura lobata</i> Needham, 1930	China	Palearctic
<i>Ischnura lorentzi</i> Ris, 1913	New Guinea	Australasian
<i>Ischnura luta</i> Polhemus, Asquith & Miller, 2000	Rota (Polynesia)	Australasian
<i>Ischnura mahechai</i> Machado, 2012	Colombia	Neotropical
<i>Ischnura nursei</i> Morton, 1907	Asia	Palearctic
<i>Ischnura ordosi</i> Bartenev, 1912	Asia	Palearctic
<i>Ischnura oreadum</i> Lieftinck, 1949	New Guinea	Australasian

Species	Distribution range	Biogeographic realms
<i>Ischnura pamela</i> Vick & Davies, 1988	New Caledonia	Australasian
<i>Ischnura patricia</i> Fraser, 1924	India	Indomalayan
<i>Ischnura perparva</i> McLachlan in Selys, 1876	W North America	Nearctic
<i>Ischnura posita</i> (Hagen, 1861)	E North America; introduced to Hawaii	Nearctic
<i>Ischnura prognata</i> (Hagen, 1861)	E North America	Nearctic
<i>Ischnura pruinescens</i> (Tillyard, 1906)	N Australia	Australasian
<i>Ischnura pumilio</i> (Charpentier, 1825)	Europe, Azores, Madeira	Palearctic
<i>Ischnura ramburii</i> (Selys in Sagra, 1857)	North and Central America; introduced to Hawaii	Neotropical- Nearctic
<i>Ischnura rhodosoma</i> Lieftinck, 1959	New Guinea	Australasian
<i>Ischnura rubella</i> Navás, 1934	Asia	Palearctic
<i>Ischnura rubilio</i> Selys, 1876	Asia (India, Bhutan)	Indomalayan
<i>Ischnura rufostigma</i> Selys, 1876	Asia	Palearctic
<i>Ischnura rufovittata</i> (Blanchard, 1843)	South America	Neotropical
<i>Ischnura rurutana</i> Englund & Polhemus, 2010	Austral islands (Polynesia)	Australasian
<i>Ischnura saharensis</i> Aguesse, 1958	North Africa, Canary Islands	Palearctic
<i>Ischnura sanguinostigma</i> Fraser, 1953	Samoa	Australasian
<i>Ischnura senegalensis</i> (Rambur, 1842)	Africa, Asia, Canary Islands	Afrotropical- Palearctic
<i>Ischnura sobrina</i> Schmidt, 1942	Peru	Neotropical
<i>Ischnura solitaria</i> Bota-Sierra, Velásquez-Vélez & Realpe, 2019	Colombia	Neotropical
<i>Ischnura spinicauda</i> Brauer, 1865	Polynesia	Australasian
<i>Ischnura stueberi</i> Lieftinck, 1932	New Guinea	Australasian
<i>Ischnura taitensis</i> Selys, 1876	Tahiti	Australasian
<i>Ischnura thelmae</i> Lieftinck, 1966	Rapa island (Polynesia)	Australasian
<i>Ischnura ultima</i> Ris, 1908	Argentina	Neotropical
<i>Ischnura verticalis</i> (Say, 1839)	E North America, Mexico	Nearctic
<i>Ischnura vinsoni</i> Fraser, 1949	Mauritius	Afrotropical
<i>Ischnura xanthocyane</i> Lieftinck, 1949	New Guinea	Australasian

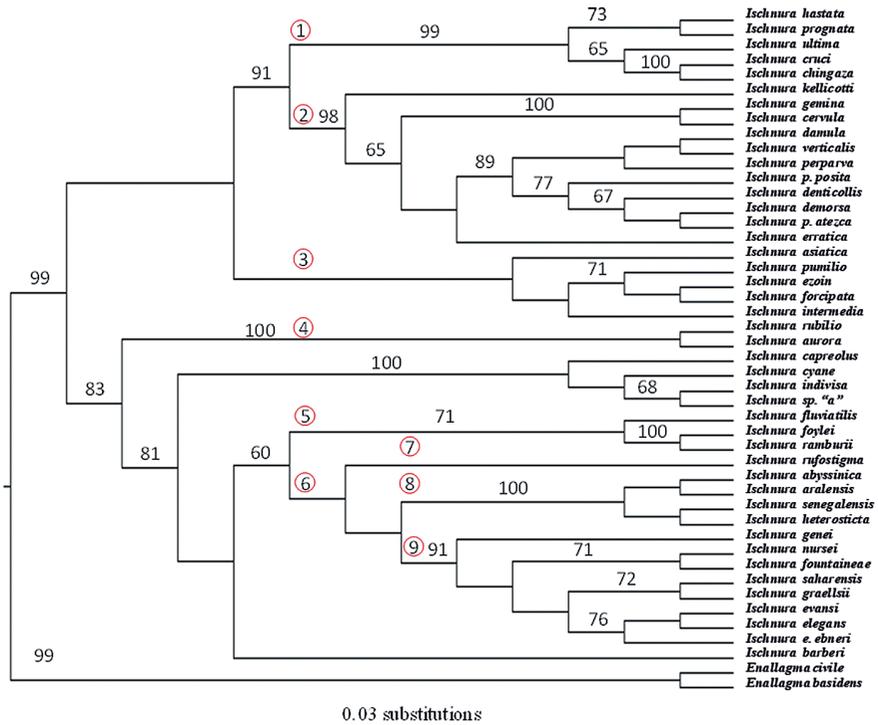
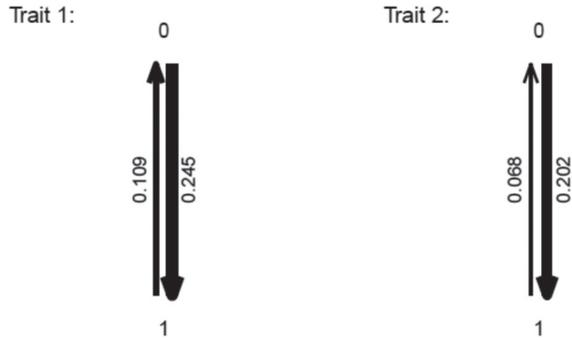


Figure S1. *Ischnura* phylogeny, including 10 novel taxa, by using maximum likelihood inference.

a) Independent model



b) Dependent model

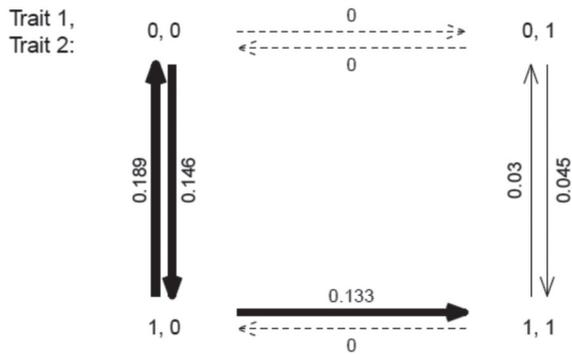
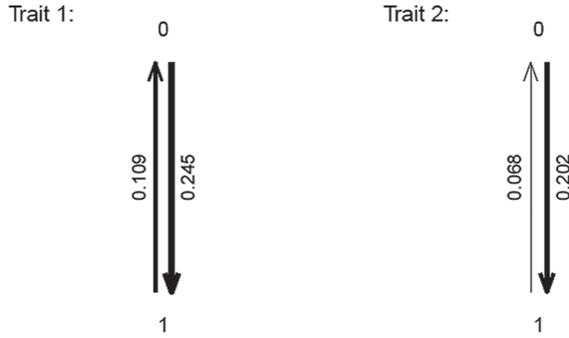


Figure S2. Correlation between female-limited colour polymorphism and mating system. Graphical representation of the transition rates between traits (females colour polymorphism and mating system) for the independent and the dependent models. Arrow widths are scaled to their average transition rates in that state.

a) Independent model



b) Dependent model

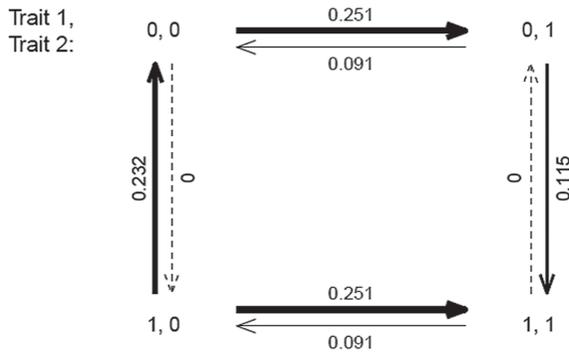


Figure S3. Graphical representation of the transition rates between traits (female colour polymorphism and mating system) for the independent and the dependent models. Arrow widths are scaled to their average transition rates in that state (female colour morphs depend on mating system).

a) Independent model



b) Dependent model

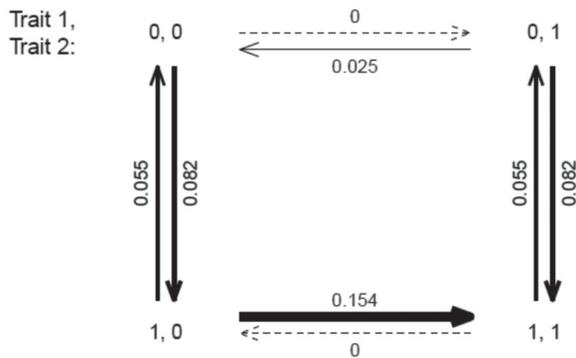


Figure S4. Mating system depends on female-limited colour polymorphism. Graphical representation of the transition rates between traits (females colour polymorphism and mating system) for the independent and the dependent models. Arrow widths are scaled to their average transition rates in that state.